

FUKUGISIDE, THE FIRST BIFLAVONOID GLYCOSIDE FROM GARCINIA SPICATA HOOK. f.

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Garcinia spicata Hook. f. (Guttiferae)--Japanese name: fukugi-- is an ever-green tree commonly found in Southeast Asia, especially in Ryukyu Islands where its bark has been used as a raw material of yellow dye. In the previous paper,<sup>1)</sup> we reported the structures of ( $\pm$ )-fukugetin (I), (+)-fukugetin (II) and ( $\pm$ )-3''-O-methyl fukugetin (III) which were isolated from the bark of Garcinia spicata. We further studied on the constituents of fresh bark of this plant and isolated a new biflavonoid glycoside, fukugiside, which is the first example of the glycoside of biflavonoid compound. This communication deals with the structural elucidation of fukugiside (IV).

Fukugiside (IV),  $C_{36}H_{30}O_{16} \cdot 2H_2O$ , m.p. 242-243° (decomp.), fine yellow needles from acetone-benzene (2 : 1), showed the following ORD data: (c = 0.31, MeOH)  $[\alpha]^{15}(\text{m}\mu) + 116 (650), + 155 (589), + 400 (450) \text{ and } + 1258 (398)$ . It gave brownish-green colour with  $FeCl_3$  and deep red colour with  $Mg-HCl$ . The spot test on a filter paper and polyamide thin-layer chromatography plate gave fine yellow colour with 5% potassium hydroxide and gave bright yellow fluorescence under the ultraviolet light. It UV absorption maxima in ethanol at 225 (shoulder), 255 (shoulder), 274, 292 and 336m $\mu$  were very similar to those of I and the maxima showed the characteristic bathochromic shift of 5,7-dihydroxy and 3',4'-ortho-dihydroxy systems<sup>2,3)</sup> on addition of sodium acetate,  $AlCl_3$  and boric acid-sodium acetate. The IR absorption (nujol mull) appears at 3250 (hydroxyl groups), 1640 (conjugated  $\delta$ -pyron), 1600 and 1570  $cm^{-1}$  (benzene rings).

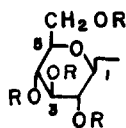
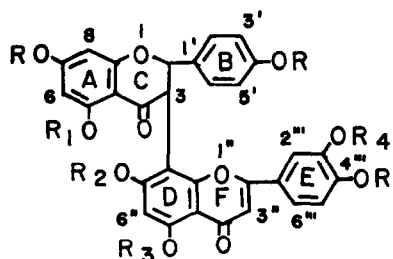
Hydrolysis of IV with sulfuric acid or cation-exchange resin (Amberlite IR-120, H-form)<sup>4)</sup> afforded, along with D-glucose, a yellow crystalline compound, which was identified with ( $\pm$ )-fukugetin (I) (mixed m.p., IR and NMR spectra). The methyl ether of this compound was also identified with an authentic sample of

heptamethyl fukugetin (V). D-Glucose was identified by paper (PPC) and thin-layer chromatography (TLC) (microcrystalline cellulose powder) and PPC, TLC and m.p. of its osazone. On acetylation IV gave a decaacetate (VI), which was negative for the ferric chloride test. NMR spectrum (60 mc. in  $\text{CDCl}_3$ ,  $\tau$  scale) of VI revealed the presence of ten acetoxyl groups (7.55 to 8.12) and C-1 to C-6 protons of the glucose moiety (multiplets, 4.50 to 6.50). The NMR spectrum (acetone- $d_6$ ) of IV showed several signals in the region between 2.40 and 7.0 (2.0 and 6.3 in pyridine- $d_5$ ), two hydrogen-bonded hydroxyl groups showed broad singlets at -3.03 and -2.13, other hydroxyl groups showed broad signals at 0.95, and all the glucose protons showed broad signals at 4.7 - 6.6 (4.3 - 6.2 in pyridine- $d_5$ ).

In order to determine the position of glucose in the molecule, IV was methylated with methyl iodide and silver oxide in dimethylformamide (Kuhn's method)<sup>5)</sup> at room temperature to give permethyl ether (VII), colourless syrup, which was negative for the ferric chloride test. The acid hydrolysis of VII yielded crystalline compound (VIII), m.p. 218-219°,  $\text{C}_{36}\text{H}_{32}\text{O}_{11}$ , which was positive for the ferric chloride test. Its UV spectrum was identical with that of V and the maxima showed the characteristic bathochromic shift of 7-hydroxy system on addition of sodium acetate. Acetylation of VIII with acetic anhydride and pyridine gave a monoacetate (IX), which was negative for the ferric chloride test. NMR spectrum of IX showed the presence of one acetoxyl group (7.82).

The NMR spectrum (acetone- $d_6$ ) of VIII showed signals due to six methoxyl groups between 6.05 and 6.35 and a pair of doublets ( $J=12$  cps) at 4.01 and 5.05 due to C-2 and C-3 protons. Protons of ring E gave rise to double doublets at 2.51 ( $J=2.5$ , 8.5 cps), a doublet ( $J=2.5$  cps) at 2.64 and a doublet ( $J=8.5$  cps) at 3.05. The C-2' and C-6' protons, and C-3' and C-5' protons gave rise to two doublets ( $J=9$  cps) at 2.75 and 3.31, respectively. The remaining three aromatic protons showed signals in the region of 3.53 - 3.84 of the spectrum. Two of them appeared as meta-coupled doublets ( $J=2.5$  cps) at 3.70 and 3.79, which were assigned to C-8 and C-6 protons respectively, whereas another proton showed a singlet at 3.55 due to C-6" proton. A singlet at 3.51 can be assigned to C-3" proton. From these data VIII was suggested to be a 5,7,4',5",3"',4"'-hexa-O-methyl fukugetin. Additional evidence for this suggestion is provided by the

NMR spectra of V, VII, VIII, 5,7,4',7'',3'',4'' -hexa-O-methyl fukugetin (X), 7,4',5'',7'',3'',4'' -hexa-O-methyl fukugetin (XI) and 7,4',7'',3'',4'' -penta-O-methyl fukugetin (XII)\*. Signals due to C-8 and C-6 protons of ring A appeared as doublets ( $J = 2.5$  cps) at 3.69 and 3.78 in V, at 3.70 and 3.79 in VII and VIII, at 3.68 and 3.77 in X, at 3.85 (singlet, 3H: C-8, C-6 and C-6'') in XI, and at 3.88 (singlet, 3H: C-8, C-6 and C-6'') in XII. The corresponding signals in V, VII, VIII and X are very similar to each other and may be assigned to C-8 and C-6 protons of ring A, respectively. The NMR spectrum is in agreement with the structure VIII. NMR spectrum (acetone- $d_6$ ) of VII showed signals due to ten methoxyl groups between 6.08 and 6.60 and multiplets at 4.92 and 5.25 - 6.68 due to C-1 and C-2 to C-6 protons of glucose. Aromatic and aliphatic protons were identical with those of heptamethyl fukugetin (V). Hydrolysis of IX with boiling ethanolic potassium hydroxide gave veratric acid and anisaldehyde.



$\beta$ -D-glc (R = H, Ac, Me)

I, II	: R = R <sub>1</sub> = R <sub>2</sub> = R <sub>3</sub> = R <sub>4</sub> = H
III	: R = R <sub>1</sub> = R <sub>2</sub> = R <sub>3</sub> = H, R <sub>4</sub> = Me
IV	: R = R <sub>1</sub> = R <sub>3</sub> = R <sub>4</sub> = H, R <sub>2</sub> = $\beta$ -D-glc
V	: R = R <sub>1</sub> = R <sub>2</sub> = R <sub>3</sub> = R <sub>4</sub> = Me
VI	: R = R <sub>1</sub> = R <sub>3</sub> = R <sub>4</sub> = Ac, R <sub>2</sub> = $\beta$ -D-glc(Ac)
VII	: R = R <sub>1</sub> = R <sub>3</sub> = R <sub>4</sub> = Me, R <sub>2</sub> = $\beta$ -D-glc(Me)
VIII	: R = R <sub>1</sub> = R <sub>3</sub> = R <sub>4</sub> = Me, R <sub>2</sub> = H
IX	: R = R <sub>1</sub> = R <sub>3</sub> = R <sub>4</sub> = Me, R <sub>2</sub> = Ac
X	: R = R <sub>1</sub> = R <sub>2</sub> = R <sub>4</sub> = Me, R <sub>3</sub> = H
XI	: R = R <sub>2</sub> = R <sub>3</sub> = R <sub>4</sub> = Me, R <sub>1</sub> = H
XII	: R = R <sub>2</sub> = R <sub>4</sub> = Me, R <sub>1</sub> = R <sub>3</sub> = H

The enzymatic hydrolysis of IV with emulsin (Sigma Chemical Co.) gave II and D-glucose to prove the  $\beta$ -linkage between glucose moiety and aglycone. From these data we propose for fukugiside the structure IV (=fukugetin-7''- $\beta$ -glucoside).

\* Methylation of (+)-fukugetin (II) with dimethyl-sulfate and potassium hydroxide afforded V, m.p. 209-210°, X, m.p. 230-231°, XI, m.p. 181-182°, and XII, m.p. 149-150°.

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